WHITE PAPER No. 69

CAR T-Cell Research: Current Clinical Challenges and Outlook

Chimeric Antigen Receptor (CAR) T-cell therapy has revolutionized the field of oncology, producing exceptionally effective and durable clinical responses for some cancer types.

CARs are recombinant receptors engineered to redirect the specificity and function of immune cells. In CAR T-cell therapy, T-cells are genetically modified to express a CAR using genetic engineering tools such as viral transduction and CRISPR-Cas9. These modified T-cells target and bind to extracellular surface cancer antigens resulting in major T-cell activation and in turn, elimination of malignant cells through processes such as induced apoptosis and cytokine release¹.

The buzz surrounding CAR T-cell therapy has translated to a steep rise in the number of associated research papers, clinical trials, and approved drugs over the past decade. Nevertheless, major limitations to CAR T therapies prevent their widespread use in oncology treatment.

Challenges associated with CAR T therapy starting materials

All current approved CAR T-cell therapies and the majority of published clinical trials have used autologous materials, which means the cells are derived from the patient for whom the therapy is being developed. However, therapies based on autologous T-cells are associated with several limitations. One major challenge, for example, is a dependence on the functional fitness of patient T-cells. And this is often diminished by the disease or prior therapies such as lymphodepleting chemotherapy and radiotherapy, which can make it difficult to obtain enough patient-derived, viable T-cells for development.

As an alternative, allogenic starting materials obtained from healthy donors yield high amounts of fully functional cells that can be used to generate multiple CAR T-cell products. While facilitating an 'off-the-shelf' therapy option, allogenic materials come with their own unique set of challenges.



Here we review the main challenges, outline the research efforts underway to overcome them, and discuss the future direction of CAR T therapies.

The main limitation of allogenic CAR T-cell products is graft-versus-host disease (GvHD). This leads to host allorejection, which ultimately hinders anti-tumor activity and can potentially result in fatalities. GvHD arises when there is a mismatch between the donor and recipient human leukocyte antigens (HLA)². In an attempt to overcome this issue, studies have worked on eliminating expression of T-cell receptors (TCR) in CAR T-cells using a range of gene editing tools; including Zinc Finger Nucleases (ZFN)³⁻⁵, Transcription Activator-Like Effector Nucleases (TALEN)^{6,7}, and CRISPR-Cas9⁸⁻¹⁰. Although high gene-editing efficiencies have been achieved using these techniques, some TCRbearing T-cells may persist that still have the potential to cause GvHD¹¹. Alternative approaches to reduce the risk of GvHD and allorejection include the elimination of HLA molecules¹², and tumor site-specific activation strategies such as hypoxia-activated CAR T-cells¹³.

WHITE PAPER | No. 69 | Page 2



Fig. 1: Simplified workflow for the production of iPSC-derived T-lymphocytes. Briefly, peripheral blood mononuclear cells (PBMCs) are recovered from a healthy donor by apheresis. Culturing PBMCs with pluripotent factors yields iPSCs which can subsequently be engineered and differentiated to yield iPSC-derived T lymphocytes.

A general limitation to both autologous and allogenic CAR T therapies is the exhaustion of T-cell ability during their amplification – which is an essential step of CAR T therapy development¹⁴. Additionally, T-cells are notoriously difficult to edit, limiting their use with engineered antigen receptors¹⁴. T-cells differentiated from either hematopoietic stem and progenitor cells (HSPCs) and induced pluripotent stem cells (iPSCs) offer an opportunity to overcome these limitations (Fig. 1). Additionally, both HSPCs and iPSCs provide rapid access to unlimited, phenotypically defined, expandable and functional T-cells. This possibility is generating increasing interest in the use of HSPCs/iPSCs for CAR T therapies¹⁵. Engineered iPSC-derived T-cells expressing CARs have been generated, demonstrated therapeutic potential in model organisms¹⁶, and recently, moved into Phase 1 clinical trials¹⁷.

Lack of Efficacy for Treating Solid Tumors

The inability of CAR T-cells to traffic to disease sites, overcome physical barriers (e.g. tumor stroma) to infiltrate solid tumors, and maintain function in a hostile tumor microenvironment has so far limited their therapeutic potential to blood disorders. Indeed, all the current approved CAR T therapies are used to treat hematological malignancies.

One strategy to widen the scope of CAR T therapies is through engineering the expression of various proteins on CAR T-cells. For instance, CAR T-cells engineered to express heparanase, an enzyme that degrades the primary component of tumor stroma (heparin sulfate proteoglycan – HSPG), have been investigated as a method to improve tumor penetration¹⁸. Certain chemokines that are known to correlate with disease burden and prognosis in multiple solid tumors have recently been exploited to improve CAR T-cell trafficking. By engineering CAR T-cells to express chemokine receptors that recognize and respond to tumor-specific chemokines, various studies have shown enhanced trafficking and significantly improved antitumor efficacy both *in silico* and *in vitro*¹⁹⁻²². Whether these results translate *in vivo* is yet to be explored.

Even if CAR T-cells successfully traffic and infiltrate the tumor, the hostile and immunosuppressive environment of tumors can prevent T-cells carrying out their effector function. Weak or non-existent responses, poor T-cell expansion, and short-term T-cell persistence are all regularly observed when treating solid tumors with CAR T-cell therapy²². Again, genetically engineered CAR T-cells are at the center of research efforts. For example, so-called "armored CARs" secrete stimulatory cytokines that support the survival, proliferation, and antitumor activity of CAR T-cells²³.

Alternative methods to manage the tumor microenvironment include combining CAR T-cell therapy with checkpoint blockade (CPB) agents. Through this combination therapy, CAR T-cells are able to infiltrate tumors while CPB agents reverse CAR T-cell inhibition to aid sustained T-cell persistence and function²⁴. Furthermore, the exploration of alternative antigen recognition domains to mitigate T-cell exhaustion is underway. Single-chain variable fragments (scFvs) are typically used to mediate antigen recognition but are linked to T-cell exhaustion. Engineered binding scaffolds, natural ligands, or receptors offer promising alternatives²⁵.

WHITE PAPER | No. 69 | Page 3

| CHALLENGES | SOLUTIONS |
|--|--|
| Trafficking/migration Complex human physiology make it difficult for CAR T-cells to home in and migrate onto malignant sites | Engineering CAR T-cells to express proteins that aid migration, such as adhesion molecules and/or chemokine receptors that match and respond to tumor-derived chemokines |
| Infiltration Physical barriers can prevent or limit CAR T-cell infiltration of a solid tumor | Engineering CAR T-cells that express enzymes capable of degrad- ing the tumor extracellular matrix |
| Maintenance of function Hostile and immunosuppressive environment of tumors can hin- der T-cells carrying out their effector function | Engineering of CAR T-cells to secrete stimulatory cytokines that support the survival, proliferation, and antitumor activity as well as rebalance the tumor microenvironment. Combination therapy: combining CAR T-cell therapy with check- point blockade (CPB) agents. |

Antigen Escape

Antigen escape is used to describe the partial or complete loss of target antigen expression on malignant T-cells in patients treated with CAR T therapy. It is a common mechanism of resistance that allows malignant cells to evade CAR T-cells. This phenomenon is responsible for the high rate of post-therapy relapse in patients treated with CAR-based therapies and is therefore an important focus for current research.

Engineering CAR T-cells that can recognize multiple targets is being explored as a solution to overcome antigen escape. These methods either use dual CAR constructs, tandem CARs, coadministration, or cotransduction (Fig. 2). Both dual CAR constructs and tandem CARs have shown promising efficacy in clinical trials, decreased antigen escape, and in some cases have even shown favorable safety profiles^{22,26}. Most CAR T-cells for blood disorders are engineered to recognize CD19, an antigen expressed by malignant cells in leukemia patients²⁷. However, the ability to recognize multiple targets hinges on the identification of novel antigens that are suitable targets for CAR T therapies. Moreover, suitable targets for solid tumors are scarce since antigen expression is highly heterogenous and many tumor antigens are also expressed on healthy cells, risking off target effects²⁸. There are many potential novel targets under investigation including some which offer opportunities to develop new therapeutic options. For instance, targeting antigens on cancer stem cells that drive tumor growth offers a promising anti-tumor therapy²⁹.



Fig. 3: . Strategies for engineering CAR T-cells to recognize multiple targets.

Summary

CAR T therapies are predominantly used after first-line treatment has failed. For instance, in the UK, CAR T therapies are vailable to children and young adults with B-cell acute lymphoblastic leukemia (ALL), whose first treatment has not worked³⁰. However, there is considerable interest in utilizing CAR T-cell therapy earlier in the course of treatment.

While CAR T therapies have already revolutionized the treatment of certain hematological malignancies, there are still limitations to their implementation and therapeutic effect. Currently, approved therapies rely on autologous material that is often insufficient for CAR T-cell manufacture due to the health

status of the patient. Even with successful CAR T-cell delivery, malignant T-cells can downregulate antigens due to selective pressures.

In solid tumors, getting CAR T-cells to traffic to and infiltrate solid tumors, penetrate physical tumor barriers and survive in the immunosuppressive tumor microenvironment is a significant challenge. While these challenges exist, CAR T-cell research is increasingly delivering novel strategies and potential solutions that drive forward more effective and safer therapies, as well as expanding the types of treatable malignancies.

Literature

- [1] Benmebarek, M. R. et al. Killing Mechanisms of Chimeric Antigen Receptor (CAR) T Cells. Int. J. Mol. Sci. 20, (2019).
- [2] Caldwell, K. J., Gottschalk, S. & Talleur, A. C. Allogeneic CAR Cell Therapy—More Than a Pipe Dream. Front. Immunol. 11, (2020).
- [3] Torikai, H. et al. Toward eliminating HLA class I expression to generate universal cells from allogeneic donors. Blood 122, 1341–1349 (2013).
- [4] Torikai, H. et al. A foundation for universal T-cell based immunotherapy: T cells engineered to express a CD19-specific chimeric-antigen-receptor and eliminate expression of endogenous TCR. Blood 119, 5697–5705 (2012).
- [5] Provasi, E. et al. Editing T cell specificity towards leukemia by zinc finger nucleases and lentiviral gene transfer. Nat. Med. 18, 807–815 (2012).
- [6] Rasaiyaah, J., Georgiadis, C., Preece, R., Mock, U. & Qasim, W. TCRαβ/CD3 disruption enables CD3-specific antileukemic T cell immunotherapy. JCI insight 3, (2018).
- [7] Sommer, C. et al. Preclinical Evaluation of Allogeneic CAR T Cells Targeting BCMA for the Treatment of Multiple Myeloma. Mol. Ther. 27, 1126–1138 (2019).
- [8] Eyquem, J. et al. Targeting a CAR to the TRAC locus with CRISPR/Cas9 enhances tumour rejection. Nature 543, 113–117 (2017).
- [9] Ren, J. et al. Multiplex Genome Editing to Generate Universal CAR T Cells Resistant to PD1 Inhibition. Clin. Cancer Res. 23, 2255–2266 (2017).
- [10] Georgiadis, C. et al. Long Terminal Repeat CRISPR-CAR-Coupled 'Universal' T Cells Mediate Potent Anti-leukemic Effects. Mol. Ther. 26, 1215–1227 (2018).
- [11] Sanber, K., Savani, B. & Jain, T. Graft-versus-host disease risk after chimeric antigen receptor T-cell therapy: the diametric opposition of T cells. Br. J. Haematol. 195, 660–668 (2021).
- [12] Kagoya, Y. et al. Genetic Ablation of HLA Class I, Class II, and the T-cell Receptor Enables Allogeneic T Cells to Be Used for Adoptive T-cell Therapy. Cancer Immunol. Res. 8, 926–936 (2020).
- [13] Juillerat, A. et al. An oxygen sensitive self-decision making engineered CAR T-cell. Sci. Rep. 7, (2017).
- [14] Flippe, L. et al. Rapid and Reproducible Differentiation of Hematopoietic and T Cell Progenitors From Pluripotent Stem Cells. Front. Cell Dev. Biol. 8, 1103 (2020).
- [15] Sadeqi Nezhad, M., Abdollahpour-Alitappeh, M., Rezaei, B., Yazdanifar, M. & Seifalian, A. M. Induced Pluripotent Stem Cells (iPSCs) Provide a Potentially Unlimited T Cell Source for CAR T-Cell Development and Off-the-Shelf Products. Pharm. Res. 38, (2021).
- [16] Iriguchi, S. et al. A clinically applicable and scalable method to regenerate T-cells from iPSCs for off-the-shelf T-cell immunotherapy. Nat. Commun. 2021 121 12, 1–15 (2021).

WHITE PAPER | No. 69 | Page 5

- [17] Global Newswire. Fate Therapeutics Announces Treatment of First Patient in Landmark Phase 1 Clinical Trial of FT819, the First-ever iPSC-derived CAR T-Cell Therapy. https://www.globenewswire.com/en/news-relea se/2021/08/02/2273241/24675/en/Fate-Therapeutics-Announces-Treatment-of-First-Patient-in-Landmark-Phase-1-Clinical-Trial-of-FT819-the-First-ever-iPSC-derived-CAR-T-Cell-Therapy.html (2021).
- [18] Caruana, I. et al. Heparanase promotes tumor infiltration and antitumor activity of CAR-redirected T lymphocytes. Nat. Med. 21, 524–529 (2015).
- [19] Whilding, L. M. et al. CAR T-Cells Targeting the Integrin αvβ6 and Co-Expressing the Chemokine Receptor CXCR2 Demonstrate Enhanced Homing and Efficacy against Several Solid Malignancies. Cancers 2019, Vol. 11, Page 674 11, 674 (2019).
- [20] Liu, G. et al. CXCR2-modified CAR T-cells have enhanced trafficking ability that improves treatment of hepatocellular carcinoma. Eur. J. Immunol. 50, 712–724 (2020).
- [21] Jin, L. et al. CXCR1- or CXCR2-modified CAR T cells co-opt IL-8 for maximal antitumor efficacy in solid tumors. Nat. Commun. 10, (2019).
- [22] Sterner, R. C. & Sterner, R. M. CAR T-cell therapy: current limitations and potential strategies. Blood Cancer J. 2021 114 11, 1–11 (2021).
- [23] Hawkins, E. R., D'souza, R. R. & Klampatsa, A. Armored CAR T-Cells: The Next Chapter in T-Cell Cancer Immunotherapy. Biologics 15, 95–105 (2021).
- [24] Grosser, R., Cherkassky, L., Chintala, N. & Adusumilli, P. S. Combination Immunotherapy with CAR T Cells and Checkpoint Blockade for the Treatment of Solid Tumors. Cancer Cell 36, 471–482 (2019).
- [25] Zajc, C. U. et al. Driving CARs with alternative navigation tools the potential of engineered binding scaffolds. FEBS J. 288, 2103–2118 (2021).
- [26] Lin, Q., Zhao, J., Song, Y. & Liu, D. Recent updates on CAR T clinical trials for multiple myeloma. Mol. Cancer 18, (2019).
- [27] Barros, L. R. C. In Search of an Ideal CAR T-Cell Antigen Target. Crit. Rev. Immunol. 41, 69–76 (2021).
- [28] Abbott, R. C., Cross, R. S. & Jenkins, M. R. Finding the Keys to the CAR: Identifying Novel Target Antigens for T Cell Redirection Immunotherapies. Int. J. Mol. Sci. 21, (2020).
- [29] Ponterio, E., De Maria, R. & Haas, T. L. Identification of Targets to Redirect CAR T Cells in Glioblastoma and Colorectal Cancer: An Arduous Venture. Front. Immunol. 11, 2339 (2020).
- [30] NHS England » CAR T-Therapy. https://www.england.nhs.uk/cancer/cdf/car-t-therapy/.

About Eppendorf

Eppendorf is a leading life science company that develops and sells instruments, consumables, and services for liquid-, sample-, and cell handling in laboratories worldwide. Its product range includes pipettes and automated pipetting systems, dispensers, centrifuges, mixers, spectrometers, and DNA amplification equipment as well as ultra-low temperature freezers, fermentors, bioreactors, CO₂ incubators, shakers, and cell manipulation systems. Consumables such as pipette tips, test tubes, microtiter plates, and single-use bioreactor vessels complement the range of highest-quality premium products.

Eppendorf was founded in Hamburg, Germany in 1945 and has more than 4,500 employees worldwide. The company has subsidiaries in 28 countries and is represented in all other markets by distributors.

Your local distributor: www.eppendorf.com/contact

 $\label{eq:spendorf} E \mbox{-} Barkhausenweg \ 1 \mbox{-} 22339 \ Hamburg \mbox{-} Germany \\ eppendorf@eppendorf.com \mbox{-} www.eppendorf.com \\ \end{tabular}$

www.eppendorf.com

Eppendorf SE reserves the right to modify its products and services at any time. This White Paper is subject to change without notice. Although prepared to ensure accuracy, Eppendorf SE assumes no liability for errors, or for any damages resulting from the application or use of this information. Viewing White Papers alone cannot as such provide for or replace reading and respecting the current version of the operating manual. Eppendorf¹ and the Eppendorf¹ and the Eppendorf¹ Bartered trademarks of Eppendorf SE, Germany. All rights reserved, including graphics and images. Copyright © 2022 by Eppendorf SE.